

DRI-CHEM 7000 Veterinary Chemistry Analyzer



DRI-CHEM® 7000 Veterinary Chemistry Analyzer Supplies

NOTE: For access to the full user manual for this product, please visit www.heska.com/productmanuals.

Remove testing supplies from refrigerator and warm to room temperature:

- Slides: 5 minutes in foil pouch.

NOTE: Remove only those needed for ordered tests.

- Reconstituted HESKA® Chemistry Control: 5 minutes.
- Electrolyte Reference Fluid: 5 minutes.

NOTE: Fill 0.5 ml or 1.5 ml Red Top sample tube depending on the REF setting in Mode 84, then return remaining RE Fluid to refrigerator.

Replacing Consumables

1. First ensure [Warming up], [Ready] or [Spot interrupted] is displayed on LCD.
2. To stop sampling, press **STOP**.

Setting Tips

1. Press **TIP/CUP** to turn sample carousel.
2. Once carousel stops, remove tip racks.
3. Fill tip racks and replace (Figure 1).
4. Press **C** to reset tip position.
5. Press **TIP/CUP** when done OR **START** to initiate testing.





quick steps guide

DRI-CHEM 7000

Veterinary Chemistry Analyzer

Replacing Consumables (Cont'd)

Replacing Mixing Cups

1. Press **TIP/CUP** to turn sample carousel.
2. Press **ENTER**.
3. Remove used mixing cups from carousel and replace new mixing cups.
4. Select <R, L or ALL> by using **SCROLL** key (<, >) and press **ENTER**.
5. Press **STOP** to complete replacement of mixing cups.
6. Press **TIP/CUP** to end replacement OR **START** to initiate testing.

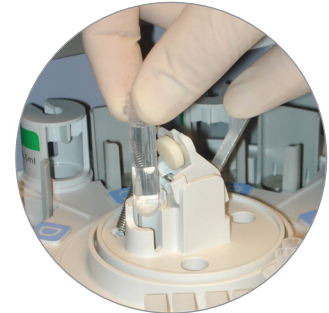
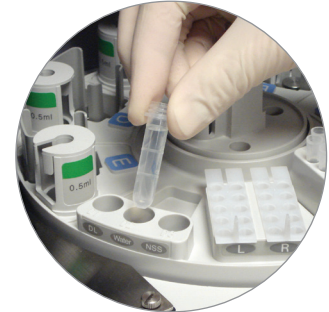
Replacing Diluent

1. Pour 1.5-ml of distilled water into a 1.5-ml Red Top non-heparinized sample tube.
2. Press **TIP/CUP** to turn sample carousel.
3. Place diluent tube in appropriate location (D2) labeled "water".
4. Press **TIP/CUP** when done OR **START** to initiate testing.

Replacing Reference Fluid

Pipette Reference Fluid into a 0.5 ml or 1.5 ml red top non-heparinized sample tube depending on the REF setting in Mode 84.

IMPORTANT: Replace RE fluid after 12 hours.



Supplies

Sample Collection and Handling

NOTE: Handle all samples and used materials as infectious waste. Wear gloves, glasses and other protective gear for your safety.

Heska-Supplied Green Top (lithium heparin) Tubes

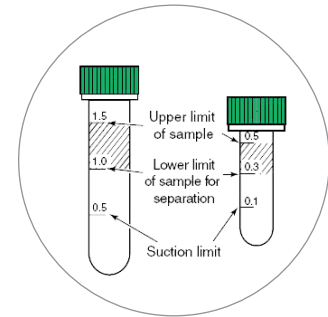
NOTE: Use with Heska-supplied centrifuge to save a step.

1. Draw patient sample using a 23-gauge or larger needle.
2. Remove needle from syringe to fill a 0.5-ml or 1.5-ml Green Top tube.

NOTE: Full 0.5-ml Green Top tube—6-test profile plus electrolytes.

Full 1.5-ml Green Top tube—Adequate volume for all available tests.

3. Fill sample tube.
 - 0.5-ml tube: Fill with at least 0.3 ml of sample (second line); Immediately invert the tube 5 to 6 times to ensure proper mixing with the lithium heparin.
 - 1.5-ml tube: Fill with at least 1 ml of sample (second line); use standard inversion or rolling to mix.
4. Spin sample in Heska-provided centrifuge for 3 to 5 minutes.



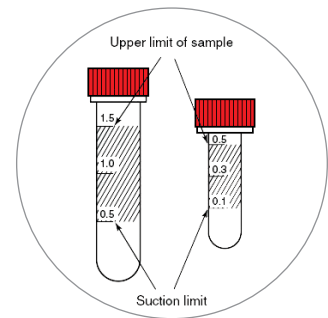
Standard Green Top Lithium Heparin or Red Top Plain Tubes for Harvesting Serum or Plasma

1. Collect sample and separate serum or plasma as you normally would.

NOTE: Red Top tube serum samples: Allow 20 minutes to clot prior to centrifugation (45 minutes for equine blood).

2. Transfer plasma or serum to a 0.5-ml Heska Red Top tube for analysis.

NOTE: Full 0.5-ml Red Top tube—adequate volume for all available tests. The 1.5-ml Red Top tube should only be used if it can be filled to at least the 1-ml mark (second line).



Basic Operation

Make sure that [Ready] or [Spot interrupted] is displayed before loading samples and slides. To stop sampling, press **STOP**.

1. Open sampler cover.
2. Place slides to be tested, barcode surface down, into slide cartridge and place weight on top.
NOTE: Handle slides by outside edges only and if using an e-wrap panel, remove the clear plastic wrap.
3. Place slide cartridge properly in appropriate sample position.
4. Remove cap from sample tube and place into appropriate sample rack. Place RE fluid if testing electrolytes. (See replacing RE Fluid section).
5. Make sure sample type on LCD is set to serum/plasma (P) using **SAMPLE** key.
6. Close sampler cover.
7. Press appropriate **SAMPLE** key (A, B, C, D or E).
NOTE: Sample indicator light will blink green.
8. Press **ID**.
9. Input sample ID (Example: 100). Press **ENTER** to accept. ① → ① → ① → ENTER
NOTE: To enter letters, press **ABC**.
10. Select appropriate species using the (▼▲) arrow keys. Press **ENTER** to accept, then press **SAVE**.
NOTE: Sample indicator light turns solid green.
11. Press **START**. Sample indicator light blinks orange during sampling.

NOTE: See next page for how to run a sample.







Running Auto-Dilution Samples

Example: To rerun Sample C with 5 times dilution:

1. Replace slides in slide cartridge C and place slide cartridge in position.
2. Press **SAMPLE KEY C**, then **RERUN**.
3. Press **DIL** to scroll to dilution factor as follows: [2] => [3] => [4] => [5]
4. Press **SAVE** to accept.
5. Press **START** to begin testing.

Running STAT Samples

Use this feature to prioritize a critical sample if more than one sample is already in progress. (Example: running STAT sample in position D.)

1. Place slides and slide weight in an unused slide cartridge.
2. Press **STAT**. LCD screen shows **[Going to stop sampling]** then **[Spot interrupted]**. Open cover and load slide cartridge and sample in open position. (Example: position D)
3. Press **SAMPLE KEY D**, then **ID** and input appropriate patient ID. Press **ENTER**.
4. Input appropriate reference range using the (▼▲) arrow keys.
5. Press **ENTER** and then **SAVE**. Sample indicator light for STAT sample turns red. 
6. Press **START** to begin testing. 

NOTE: Original sample that was interrupted will finish first. STAT sample will be processed next, followed by any remaining programmed samples.

Interrupting Samples

Press **STOP** key once to interrupt or stop sampling, or twice to cancel test programming.

NOTE: Do not open waste disposal drawer during sample processing.



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Veterinary Chemistry Analyzer

Calibration

This step can be done during warm-up [WARMING UP] or when in [READY] mode.

- For new slide lot #, insert calibration card into top of card reader on right side of screen and pull down.

NOTE: Test name and lot #'s will appear on LCD screen. Analyzer will store 6 lots per analyte or panel.

Controls

NOTE: Always wear gloves when running controls.

1. Run HESKA Chemistry Control solution as you would a patient sample by transferring into a 0.5-ml Red Top tube.

Daily Maintenance

1. Keep lid closed except when loading and unloading slides.
2. Empty waste disposal drawer.
3. Change reference fluid every 12 hours.
4. Remove sample tubes when finished testing.
5. Clean any spills on the spotting slide transit area.



DRI-CHEM 7000 Veterinary Chemistry Analyzer Slides		Quantity Per Box
CAT 6330-ALB	Albumin	24/box
CAT 6330-ALP	Alkaline Phosphatase	24/box
CAT 6330-ALT	ALT (GPT)	24/box
CAT 6330-AMY	Amylase	24/box
CAT 6330-AST	AST (GOT)	24/box
CAT 6330-BUN	BUN	24/box
CAT 6330-CA	Calcium	24/box
CAT 6330-CHOL	Total Cholesterol	24/box
CAT 6330-CK	CK	24/box
CAT 6330-CREA	Creatinine	24/box
CAT 6330-GGT	GGT	24/box
CAT 6330-GLU	Glucose	24/box
CAT 6330-LDH	Lactate Dehydrogenase	24/box
CAT 6330-LIP	Lipase	24/box
CAT 6330-MG	Magnesium	24/box
CAT 6330-NH ₃	Ammonia	24/box + NH ₃ control
CAT 6330-PHOS	Phosphorous	24/box
CAT 6330-TBILI	Total Bilirubin	24/box
CAT 6330-TP	Total Protein	24/box
CAT 6330-TRIG	Triglycerides	24/box
CAT 6330-UA	Uric Acid	24/box
CAT 6330-LYTES	Electrolytes	24/box
CAT 6330-PRESURG/EWRAP	Pre-Surgical Panel (ALP, ALT, BUN, CREA, GLU, TP)	72 slides/box (12 panels/box)
CAT 6330-EQ	Equine Panel (ALB, AST, BUN, CA, CK, CREA, GGT, GLU, LDH, PHOS, TP, T-BILI)	24 slides/box (2 panels/box)
CAT 6330-KIDNEY	Kidney Panel (ALB, BUN, CA, CREA, PHOS, TP)	24 slides/box (4 panels/box)
CAT 6330-LIVER	Liver Panel (ALB, ALP, ALT, GGT, GLU, T-BILI)	24 slides/box (4 panels/box)
CAT 6330-COMP/EWRAP	Comprehensive Panel (ALP, ALT, BUN, CREA, GLU, TP, T-BILI, ALB, PHOS, CA, CHOL, GGT)	72 slides/box (6 panels/box)
CAT 6330-PLUS	Plus Panel (LIP, AMY, MG, TRIG, AST, LYTES)	24 slides/box (4 panels/box)



For further assistance, please call Heska's Technical Support Services at 800.464.3752, option 3.